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I

(Acts adopted under the EC Treaty/Euratom Treaty whose publication is obligatory)

REGULATIONS

COMMISSION REGULATION (EC) No 120/2008**of 11 February 2008****establishing the standard import values for determining the entry price of certain fruit and vegetables**

THE COMMISSION OF THE EUROPEAN COMMUNITIES,

Having regard to the Treaty establishing the European Community,

Having regard to Commission Regulation (EC) No 1580/2007 of 21 December 2007 laying down implementing rules of Council Regulations (EC) No 2200/96, (EC) No 2201/96 and (EC) No 1182/2007 in the fruit and vegetable sector ⁽¹⁾, and in particular Article 138(1) thereof,

Whereas:

- (1) Regulation (EC) No 1580/2007 lays down, pursuant to the outcome of the Uruguay Round multilateral trade negotiations, the criteria whereby the Commission fixes

the standard values for imports from third countries, in respect of the products and periods stipulated in the Annex thereto.

- (2) In compliance with the above criteria, the standard import values must be fixed at the levels set out in the Annex to this Regulation,

HAS ADOPTED THIS REGULATION:

Article 1

The standard import values referred to in Article 138 of Regulation (EC) No 1580/2007 shall be fixed as indicated in the Annex hereto.

Article 2

This Regulation shall enter into force on 12 February 2008.

This Regulation shall be binding in its entirety and directly applicable in all Member States.

Done at Brussels, 11 February 2008.

For the Commission

Jean-Luc DEMARTY

*Director-General for Agriculture and
Rural Development*

⁽¹⁾ OJ L 350, 31.12.2007, p. 1.

ANNEX

to Commission Regulation of 11 February 2008 establishing the standard import values for determining the entry price of certain fruit and vegetables

(EUR/100 kg)

CN code	Third country code ⁽¹⁾	Standard import value
0702 00 00	IL	143,2
	MA	45,8
	MK	36,8
	TN	111,3
	TR	91,9
	ZZ	85,8
0707 00 05	EG	208,2
	JO	202,1
	MA	175,9
	TR	140,0
	ZZ	181,6
0709 90 70	MA	48,0
	TR	140,3
	ZZ	94,2
0709 90 80	EG	349,4
	ZZ	349,4
0805 10 20	EG	48,2
	IL	55,1
	MA	54,0
	TN	50,2
	TR	70,8
	ZZ	55,7
0805 20 10	IL	106,6
	MA	114,6
	TR	72,2
	ZZ	97,8
0805 20 30, 0805 20 50, 0805 20 70, 0805 20 90	CN	41,9
	EG	83,3
	IL	73,1
	JM	97,3
	MA	131,7
	TR	80,4
	ZZ	84,6
0805 50 10	EG	78,7
	IL	114,8
	MA	77,4
	TR	123,0
	ZZ	98,5
0808 10 80	CA	102,8
	CN	88,7
	MK	40,9
	US	114,4
	ZZ	86,7
0808 20 50	CN	68,2
	US	119,3
	ZA	106,6
	ZZ	98,0

⁽¹⁾ Country nomenclature as fixed by Commission Regulation (EC) No 1833/2006 (OJ L 354, 14.12.2006, p. 19). Code 'ZZ' stands for 'of other origin'.

COMMISSION REGULATION (EC) No 121/2008**of 11 February 2008****laying down the method of analysis for the determination of starch content in preparations of a kind used in animal feeding (CN code 2309)**

THE COMMISSION OF THE EUROPEAN COMMUNITIES,

HAS ADOPTED THIS REGULATION:

Having regard to the Treaty establishing the European Community,

Having regard to Council Regulation (EEC) No 2658/87 of 23 July 1987 on the tariff and statistical nomenclature and on the Common Customs Tariff⁽¹⁾, and in particular Article 9(1)(a) thereof,

Whereas:

- (1) In order to ensure that preparations of a kind used in animal feeding (CN 2309) receive uniform treatment on import throughout the Community, it is necessary that, when laying down analytic methods, account should be taken of the scientific and technological evolution of those methods.
- (2) In accordance with the third Commission Directive 72/199/EEC of 27 April 1972 establishing Community methods of analysis for the official control of feeding-stuffs⁽²⁾, for the determination of the starch content in preparations of a kind used in animal feeding, the polarimetric method (also called: modified Ewers method), described in point 1 of Annex I to that Directive is to be applied.
- (3) In the light of studies carried out by the experts of the customs laboratories of the Member States it is necessary to provide that where the polarimetric method laid down in Directive 72/199/EEC cannot be applied for the determination of the starch content in the mentioned preparations, an enzymatic analytical method should be applied. It is therefore appropriate to specify how that enzymatic method is to be carried out.
- (4) The measures provided for in this Regulation are in accordance with the opinion of the Tariff and Statistical Nomenclature Section of the Customs Code Committee,

Article 1

By derogation from Article 1 of Directive 72/199/EEC, the starch content by weight of preparations of a kind used in animal feeding within the meaning of CN code 2309 shall be determined by the enzymatic analytical method laid down in the Annex to this Regulation in cases where the following feed materials are present in significant amounts:

- (a) (sugar) beet products such as (sugar) beet pulp, (sugar) beet molasses, (sugar) beet pulp-molassed, (sugar) beet vinasse, (beet) sugar;
- (b) citrus pulp;
- (c) linseed; linseed expeller; linseed extracted;
- (d) rape seed; rape seed expeller; rape seed extracted; rape seed hulls;
- (e) sunflower seed; sunflower seed extracted; sunflower seed, partially decorticated, extracted;
- (f) copra expeller; copra extracted;
- (g) potato pulp;
- (h) dehydrated yeast;
- (i) products rich in inulin (e.g. chips and meal of Jerusalem artichokes);
- (j) greaves.

Article 2

This Regulation shall enter into force on the twentieth day following that of its publication in the *Official Journal of the European Union*.

⁽¹⁾ OJ L 256, 7.9.1987, p. 1. Regulation as last amended by Commission Regulation (EC) No 1352/2007 (OJ L 303, 21.11.2007, p. 3).

⁽²⁾ OJ L 123, 29.5.1972, p. 6. Directive as last amended by Directive 1999/79/EC (OJ L 209, 7.8.1999, p. 23).

This Regulation shall be binding in its entirety and directly applicable in all Member States.

Done at Brussels, 11 February 2008.

For the Commission
László KOVÁCS
Member of the Commission

ANNEX

ENZYMATIC METHOD FOR THE DETERMINATION OF THE STARCH CONTENT IN PREPARATIONS USED IN ANIMAL FEEDING USING HIGH PRESSURE LIQUID CHROMATOLOGY (HPLC)**1. Scope**

This method describes the enzymatic determination of the starch content in animal feeding. The starch content is derived from the quantitative determination of glucose after enzymatic breakdown of the present starch into glucose. All measured glucose is considered to be derived from the starch, present in the sample.

2. Definitions

In this method, the content of starch and its high molecular mass breakdown products, insoluble in 40 % ethanol, is determined. The starch content is expressed in % (m/m).

3. Principle

The samples are homogenised by milling. The sample is washed with 40 % ethanol to eliminate soluble sugars and soluble products of starch decomposition.

The enzyme thermostable alpha-amylase is added to the suspension. This enzyme breaks down the starch at 100 °C in smaller chains, whether or not the starch is fully in solution.

Large lumps of starch are broken down very slowly. Therefore it is necessary that the samples should be wholly solved or should be present in the form of a suspension containing very small solid parts.

Then the second enzyme amyloglucosidase is added, which hydrolyses the degraded glucose chains into glucose at 60 °C.

After clarification of the liquid whereby the present proteins, fats and residues are discarded after filtration, a clear solution is obtained which may be used for HPLC.

Separation of the present sugars is done by HPLC.

4. Reagents and other materials

Use reagents of recognized analytical grade and demineralized water.

4.1. Ethanol 40 % vol. in water

4.2. Glucose, min 99 %

4.3. Solution of Amyloglucosidase (1,4-alpha-D-Glucan glucohydrolase) from *Aspergillus niger* (enzyme activity > 5 000 U/ml). Storage at ca. 4 °C

Alternatively, also amyloglucosidase powder can be used.

4.4. Thermostable alpha-amylase (1,4-alpha-D-Glucan-glucohydrolase). Storage at ca. 4 °C

4.5. Zinc acetate dihydrate, p.a.

4.6. Potassium hexacyanoferrate (II) ($K_4[Fe(CN)_6 \cdot 3H_2O]$), extra pure.

4.7. Sodium acetate anhydrous, p.a.

4.8. Glacial acetic acid, 100 % (v/v)

4.9. Sodium acetate buffer (0,2 mol/l).

Weigh 16,4 gram sodium acetate (4.7) into a beaker glass. Dissolve in water and rinse into a volumetric flask of 1 000 ml. Dilute to the mark with water and adjust the pH to 4,7 with acetic acid (by use of a pH-meter (5.11)). This solution may be used for max 6 months with storage at 4 °C.

4.10. Amyloglucosidase solution (enzyme activity > 250 U/ml)

Prepare a solution of 5 ml of amyloglucosidase-solution (4.3) or 660 mg amyloglucosidase powder in a final volume of 100 ml by using sodium acetate buffer (4.9). Prepare instantaneously.

4.11. Reference solution

Prepare solutions of glucose in water, as conventionally used in the HPLC analysis.

4.12. Reagent for clarification (Carrez I)

Dissolve 219,5 gram zinc acetate (4.5) in water in a beaker glass. Rinse into a volumetric flask of 1 000 ml, add 30 ml acetic acid (4.8). Mix thoroughly and make up with water. This solution may be used for max 6 months while stored at ambient temperature.

Other clarification reagents, equivalent to Carrez solution, may be used.

4.13. Reagent for clarification (Carrez II)

Dissolve 106,0 gram potassium hexacyanoferrate (II) (4.6) in water in a beaker glass. Rinse into a volumetric flask of 1 000 ml. Mix thoroughly and make up with water. This solution may be used for max. 6 months while stored at ambient temperature.

Other clarification reagents, equivalent to Carrez solution, may be used.

4.14. Mobile phase

A mobile phase is prepared which is conventionally used in the HPLC analysis of sugars. In case of using an aminopropyl silicagel column, e.g., a common mobile phase is a mixture of HPLC graded water and acetonitrile.

5. **Apparatus**

5.1. Standard laboratory glassware

5.2. Centrifuge giving 1 000 g or more (calculated at the centre of the tube)

5.3. Glass centrifuge tubes of 100 ml

5.4. Magnetic stirrer

5.5. Magnetic rods

5.6. Fluted filters, e.g., 185 mm

5.7. Syringe filters, 0,45 µm, suitable for aqueous solutions

5.8. Sample vials suitable for the HPLC autosampler

5.9. 100 ml volumetric flasks

5.10. Plastic syringes, 5 and 10 ml

5.11. pH-meter

5.12. Water bath with thermostat, adjustable to 60 °C and 100 °C

5.13. Heating plates with magnetic stirrers

5.14. HPLC Apparatus

5.14.1. Pump, fit for zero-pulsation

5.14.2. Autosampler

5.14.3. Column and pre-column, suitable for analysis of sugars

5.14.4. Column oven, temperature range between ambient and 40 °C

5.14.5. Detector, suitable for analysis of sugars, e.g., Refractive Index Detector

5.14.6. Integration system

6. **Procedure**

6.1. General

Samples are analysed in singular.

6.2. Preparation of the sample for several types of products

The product is homogenised by milling.

6.3. Sample portion

The starch content is estimated with help from the ingredient declaration. The amount of sample (weighed to 0.1 mg accuracy) can be estimated from:

$$\text{amount of sample (g)} = \frac{\text{volume of volumetric flask (100 ml)}}{\text{estimated starch content (\%)}}$$

6.4. Blank determination

The blank is determined by performing a complete analysis (as described in 6.5), without adding sample. The result of the blank determination is used in the calculation of the starch content (7.1).

6.5. Analysis

6.5.1. Preparation of the samples

Mix the sample by shaking or stirring. The chosen test portion (6.3) is weighed into a centrifuge tube (5.3) and 50 ml 40 % ethanol (4.1) is added. Stir in a magnetic stirrer for 20 minutes at ambient temperature. Leave the magnetic rod in the tube and centrifuge for five minutes. Aspirate carefully and remove the liquid phase (e.g., with a Pasteur pipette). Repeat this extraction procedure with two times 25 ml ethanol (4.1). Transfer the residue into a volumetric flask of 100 ml (5.9) with circa 70 ml water. After dissolving or suspending, add 100 microliter of thermostable alpha-amylase (4.4) and heat at 100 °C for 1 hour e.g., in a water bath (5.12). Cool to 60 °C in a water bath, and add 5 ml of a solution of amyloglucosidase (4.10). Place flask for 30 min in a water bath at 60 °C. Cool to ambient temperature, clarify the sample by adding 1 ml Carrez I (4.12), shake and then add 1 ml Carrez II (4.13). Carrez I and II may be added before or after cooling. Dilute to the mark with water, homogenize and filter the solution through a fluted filter (5.6). Collect the sample extract.

6.5.2. Processing of the sample extracts

Filter the extracts through a disc filter (5.7) with a syringe (5.10) that has been preflushed with the extract. Collect the filtrates in vials (5.8).

Note: The disc filter can be used several times. It should be flushed with the next extract to prevent contamination of the previous extract.

6.6. Chromatography

HPLC is performed as conventionally for analysis of sugars. Because the samples are extracted with ethanol/water, glucose is the main sugar to be analysed. If the HPLC analysis shows traces of maltose, then this may indicate incomplete conversion of starch.

7. Calculation and expression of results

7.1. Calculation of the HPLC results

From the results of the HPLC analysis, the content (% m/m) of glucose is calculated. The enzymatic solution of amyloglucosidase (4.3) is stabilised with glucose. Furthermore, thermostable alpha-amylase (4.4) is stabilised with sucrose, which can partially be converted into glucose by invertase-activity of amyloglucosidase. Therefore, the glucose concentration (% m/v) measured should be corrected for the glucose concentration (% m/v) in the blank. The glucose content (% m/m) corrected for the blank is then calculated from the corrected glucose concentration, the sample weight and the calibration with reference solutions (4.11).

7.2. Calculation of the starch content

The starch content (% m/m) is calculated from the glucose content (% m/m) after correction for the blank.

$$\text{Starch content} = 0,9 * \text{Glucose corrected}$$

8. Precision

8.1. Inter-laboratory test

Details of an inter-laboratory test on the precision of the method are summarized in 8.4.

8.2. Repeatability

The absolute difference between two independent single test results, obtained using the same method on identical test material in the same laboratory by the same operator using the same equipment within a short interval of time, will in not more than 5 % of cases be greater than the repeatability limit of 1,1 % (m/m). The repeatability limit was derived from the pooled results of an inter-laboratory test (see 8.4).

8.3. Reproducibility

The absolute difference between two single test results, obtained using the same method on identical test material in different laboratories with different operators using different equipment, will in not more than 5 % of cases be greater than the reproducibility limit of 3,7 % (m/m). The reproducibility limit was derived from the pooled results of an inter-laboratory test (see 8.4).

8.4. Results of inter-laboratory test

An inter-laboratory test was carried out in 2005 and 2006 with participation of the European Customs laboratories. This test was performed in accordance with ISO 5725 and the IUPAC protocol (W. Horwitz, Pure and Applied Chemistry, vol. 67, 1995, p. 331-343). The precision data are given in the table below.

Statistical results of the inter-laboratory study

	Sample				
	1	2	3	4	5
Number of laboratories after eliminating outliers	25	26	26	25	24
Number of accepted results	50	52	52	50	48
Mean starch content (% m/m)	31,2	14,4	25,1	12,9	27,8
Repeatability standard deviation s_r (% m/m)	0,4	0,3	0,6	0,2	0,3
Repeatability limit r (% m/m)	1,1	0,8	1,7	0,7	0,9
Reproducibility standard deviation s_R (% m/m)	1,7	0,8	1,7	0,9	1,3
Reproducibility limit R (% m/m)	4,8	2,2	4,7	2,5	3,7

Samples

- 1: dry dog food
- 2: dry cat food
- 3: dry cat food (sample 2) with addition of starch
- 4: dry cat food (sample 2) with addition of beet pulp
- 5: commercial pet food

II

(Acts adopted under the EC Treaty/Euratom Treaty whose publication is not obligatory)

AGREEMENTS

COUNCIL

Information relating to the entry into force of the Protocol to the Agreement on Cooperation and Customs Union between the European Economic Community and the Republic of San Marino, regarding the participation, as contracting parties, of the Republic of Bulgaria and Romania, following their accession to the European Union ⁽¹⁾

The procedures necessary for the entry into force of the Protocol to the Agreement on Cooperation and Customs Union between the European Economic Community and the Republic of San Marino, regarding the participation, as contracting parties, of the Republic of Bulgaria and Romania, following their accession to the European Union, signed in Brussels on 20 November 2007, having been completed on 31 January 2008, this Protocol entered into force, in conformity with its Article 5, on 1 February 2008.

⁽¹⁾ OJ L 325, 11.12.2007, p. 84.

CORRIGENDA**Corrigendum to Council Regulation (EC) No 74/2008 of 20 December 2007 on the establishment of the 'ARTEMIS Joint Undertaking' to implement a Joint Technology Initiative in Embedded Computing Systems**

(Official Journal of the European Union L 30 of 4 February 2008)

On page 58, in the signature formula:

for: 'Done at Brussels, 20 December 2008.'

read: 'Done at Brussels, 20 December 2007.'
