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Acts whose titles are printed in light type are those relating to day-to-day management of agricultural matters, and are generally valid for a limited period.

The titles of all other Acts are printed in bold type and preceded by an asterisk.

II

(Acts whose publication is not obligatory)

COMMISSION

FIRST COMMISSION DIRECTIVE

of 1 March 1976

establishing Community methods of sampling for the official control of feedingstuffs

(76/371/EEC)

THE COMMISSION OF THE EUROPEAN
COMMUNITIES,

HAS ADOPTED THIS DIRECTIVE:

Article 1

Having regard to the Treaty establishing the European Economic Community,

Having regard to the Council Directive of 20 July 1970 on the introduction of Community methods of sampling and analysis for the official control of feedingstuffs ⁽¹⁾, as last amended by the Act of Accession ⁽²⁾, and in particular Article 2 thereof,

The Member States shall require that sampling for the official control of feedingstuffs, as regards the determination of constituents, additives and undesirable substances and products, with the exception of residues of pesticides and micro-organisms, be carried out in accordance with the methods described in the Annex to this Directive.

Article 2

Whereas that Directive requires that official control of feedingstuffs be carried out using Community methods of sampling and analysis for the purpose of checking compliance with requirements arising under the provisions laid down by law, regulation or administrative action concerning the quality and composition of feedingstuffs;

The Member States shall, not later than 1 January 1977, bring into force the laws, regulations or administrative provisions necessary to comply with this Directive. They shall forthwith notify the Commission thereof.

Article 3

Whereas, as a first stage, methods of sampling should be laid down for the control of the constituents of feedingstuffs and their additives and for the control of undesirable substances and products, with the exception of residues of pesticides and micro-organisms, which these feedingstuffs may contain;

This Directive is addressed to the Member States.

Done at Brussels, 1 March 1976.

For the Commission

P. J. LARDINOIS

Member of the Commission

Whereas the measures provided for in this Directive are in accordance with the opinion of the Standing Committee for Feedingstuffs,

⁽¹⁾ OJ No L 170, 3. 8. 1970, p. 2.

⁽²⁾ OJ No L 73, 27. 3. 1972, p. 14.

ANNEX

METHODS OF SAMPLING

1. PURPOSE AND SCOPE

Samples intended for the official control of feedingstuffs, for quality and composition, shall be taken according to the methods described below. Samples thus obtained shall be considered as representative of the sampled portions.

2. SAMPLING OFFICERS

The samples shall be taken by officers authorized for that purpose by the Member States.

3. DEFINITIONS

Sampled portion: A quantity of product constituting a unit, and having characteristics presumed to be uniform.

Incremental sample: A quantity taken from one point in the sampled portion.

Aggregate sample: An aggregate of incremental samples taken from the same sampled portion.

Reduced sample: A representative part of the aggregate sample, obtained from the latter by a process of reduction.

Final sample: A part of the reduced sample or of the homogenized aggregate sample.

4. APPARATUS

4.1. The sampling apparatus must be made of materials which cannot contaminate the products to be sampled. Such apparatus may be officially approved by the Member States.

4.2. Apparatus recommended for the sampling of solid feedingstuffs

4.2.1. *Manual sampling*

4.2.1.1. Flat-bottomed shovel with vertical sides.

4.2.1.2. Sampling spear with a long split or compartments. The dimensions of the sampling spear must be appropriate to the characteristics of the sampled portion (depth of container, dimensions of sack, etc.) and to the particle size of the feedingstuff.

4.2.2. *Mechanical sampling*

Approved mechanical apparatus may be used for the sampling of moving feedingstuffs.

4.2.3. *Divider*

Apparatus designed to divide the sample into approximately equal parts may be used for taking incremental samples and for the preparation of reduced and final samples.

5. QUANTITATIVE REQUIREMENTS

5.A. In relation to the control of substances or products uniformly distributed throughout the feedingstuff

5.A.1. *Sampled portion*

The size of the sampled portion must be such that each of its constituent parts can be sampled.

5.A.2. *Incremental samples*

5.A.2.1.	Loose feedingstuffs:	Minimum number of incremental samples:
5.A.2.1.1.	sampled portions not exceeding 2.5 metric tons	seven
5.A.2.1.2.	sampled portions exceeding 2.5 metric tons	$\sqrt{20 \text{ times the number of metric tons making up the sampled portion (a), up to a maximum of 40 incremental samples}}$
5.A.2.2.	Packaged feedingstuffs:	Minimum number of packages to be sampled (b):
5.A.2.2.1.	Packages of more than one kg:	
5.A.2.2.1.1.	sampled portions of one to four packages	all packages
5.A.2.2.1.2.	sampled portions of five to 16 packages	four
5.A.2.2.1.3.	sampled portions of more than 16 packages	$\sqrt{\text{number of packages making up the sampled portion (a), up to maximum of 20 packages}}$
5.A.2.2.2.	Packages not exceeding 1 kg	four
5.A.2.3.	Liquid or semi-liquid feedingstuffs:	Minimum number of containers to be sampled (b):
5.A.2.3.1.	Containers of more than one litre:	
5.A.2.3.1.1.	sampled portions of one to four containers	all containers
5.A.2.3.1.2.	sampled portions of five to 16 containers	four
5.A.2.3.1.3.	sampled portions of more than 16 containers	$\sqrt{\text{number of containers making up the sampled portion (a), up to a maximum of 20 containers}}$
5.A.2.3.2.	Containers not exceeding one litre	four
5.A.2.4.	Feed blocks and mineral licks	Minimum number of blocks or licks to be sampled (b): one block or lick per sampled portion of 25 units, up to a maximum of four blocks or licks

5.A.3. *Aggregate sample*

A single aggregate sample per sampled portion is required. The total amount in the incremental samples making up the aggregate sample shall be not less than the following:

5.A.3.1.	Loose feedingstuffs	4 kg
5.A.3.2.	Packaged feedingstuffs:	

(a) Where the number obtained is a fraction, it should be rounded up to the next whole number.

(b) For packages or containers whose contents do not exceed 1 kg or one litre and for blocks or licks weighing not more than 1 kg each, an incremental sample shall be the contents of one original package or container, one block or one lick.

5.A.3.2.1.	packages of more than 1 kg	4 kg
5.A.3.2.2.	packages not exceeding 1 kg	weight of the contents of four original packages
5.A.3.3.	Liquid or semi-liquid feedingstuffs:	
5.A.3.3.1.	containers of more than one litre	four litres
5.A.3.3.2.	containers not exceeding one litre	volume of the contents of four original containers
5.A.3.4.	Feed blocks or mineral licks:	
5.A.3.4.1.	each weighing more than 1 kg	4 kg
5.A.3.4.2.	each weighing not more than 1 kg	weight of four original blocks or licks
5.A.4.	<i>Final samples</i>	
	The aggregate sample gives the final samples on reduction when necessary. Analysis of at least one final sample is required. The amount in the final sample for analysis shall be not less than the following:	
	Solid feedingstuffs	500 g
	Liquid or semi-liquid feedingstuffs	500 ml
5.B.	In relation to the control of undesirable substances or products likely to be distributed non-uniformly throughout the feedingstuffs, such as aflatoxins, rye ergot, castor-oil plant and crotalaria in straight feedingstuffs (c)	
5.B.1.	<i>Sampled portion: see 5.A.1.</i>	
5.B.2.	<i>Incremental samples</i>	
5.B.2.1.	Loose feedingstuffs: see 5.A.2.1.	
5.B.2.2.	Packaged feedingstuffs:	Minimum number of packages to be sampled:
5.B.2.2.1.	sampled portions consisting of one to four packages	all packages
5.B.2.2.2.	sampled portions consisting of five to 16 packages	four
5.B.2.2.3.	sampled portions consisting of more than 16 packages	$\sqrt{\text{number of packages making up the sampled portion (a), up to a maximum of 40 packages}}$
5.B.3.	<i>Aggregate samples</i>	
	The number of aggregate samples will vary with the size of the sampled portion. The minimum number of aggregate samples per sampled portion is given below. The total weight of the incremental samples making up each aggregate sample shall be not less than 4 kg.	

(a) Where the number obtained is a fraction, it should be rounded up to the next whole number.

(c) The methods provided for in 5.A are for use in the control of aflatoxins, rye ergot, castor-oil plant and crotalaria in complete and supplementary feedingstuffs.

5.B.3.1. Loose feedingstuffs

Size of the sampled portion in metric tons:	Minimum number of aggregate samples per sampled portion:
up to 1	1
more than 1 and up to 10	2
more than 10 and up to 40	3
more than 40	4

5.B.3.2. Packaged feedingstuffs size of the sampled portion in number of packages:

1 to 16	1
17 to 200	2
201 to 800	3
more than 800	4

5.B.4. *Final samples*

Each aggregate sample gives the final samples on reduction. Analysis of at least one final sample *per aggregate sample* is required. The weight of the final sample for analysis may not be less than 500 g.

6. INSTRUCTIONS FOR TAKING, PREPARING AND PACKAGING THE SAMPLES

6.1. General

The samples must be taken and prepared as quickly as possible bearing in mind the precautions necessary to ensure that the product is neither changed nor contaminated. Instruments and also surfaces and containers intended to receive samples must be clean and dry.

6.2. Incremental samples

6.2.A. *In relation to the control of substances or products uniformly distributed throughout the feedingstuff*

Incremental samples must be taken *at random throughout the whole sampled portion* and they must be of approximately equal sizes.

6.2.A.1. Loose feedingstuffs

An imaginary division shall be made of the sampled portion into a number of approximately equal parts. A number of parts corresponding to the number of incremental samples required in accordance with 5.A.2 shall be selected at random and at least one sample taken from each of these parts.

Where appropriate, sampling may be carried out when the sampled portion is being moved (loading or unloading).

6.2.A.2. Packaged feedingstuffs

Having selected the required number of packages for sampling as indicated in 5.A.2, part of the contents of each package shall be removed using a spear or shovel. Where necessary, the samples shall be taken after emptying the packages separately.

6.2.A.3. Homogeneous or homogenizable liquid or semi-liquid feedingstuffs

Having selected the required number of containers for sampling as indicated in 5.A.2, the contents shall be homogenized if necessary and an amount taken from each container.

The incremental samples may be taken when the contents are being discharged.

6.2.A.4. Non-homogenizable, liquid or semi-liquid feedingstuffs

Having selected the required number of containers for sampling as indicated in 5.A.2, samples shall be taken from different levels.

Samples may also be taken when the contents are being discharged but the first fractions should be discarded:

In either case the total volume taken must not be less than 10 litres.

6.2.A.5. Feed blocks and mineral licks

Having selected the required number of blocks or licks for sampling as indicated in 5.A.2, a part of each block or lick shall be taken.

6.2.B. *In relation to the control of undesirable substances or products likely to be distributed non-uniformly throughout the feedingstuff, such as aflatoxins, rye ergot, castor-oil plant and crotalaria in straight feedingstuffs*

An imaginary division shall be made of the sampled portion into a number of approximately equal parts, *corresponding to the number of aggregate samples provided for in 5.B.3*. If this number is greater than one, the total number of incremental samples provided for in 5.B.2 shall be distributed approximately equally over the different parts. Then take samples of approximately equal sizes (d) and such that the total amount in the samples from each part is not less than the minimum 4 kg quantity required for each aggregate sample. *Incremental samples taken from different parts shall not be aggregated.*

6.3. Preparation of aggregate samples

6.3.A. *in relation to the control of substances of products distributed uniformly throughout the feedingstuff*

The incremental samples shall be mixed to form a single aggregate sample.

6.3.B. *In relation to the control of undesirable substances or products likely to be distributed non-uniformly throughout the feedingstuff, such as aflatoxins, rye ergot, castor-oil plant and crotalaria in straight feedingstuffs*

The incremental samples from each part of the sampled portion shall be mixed and the number of aggregate samples provided for in 5.B.3, made up *taking care to note the origin of each aggregate sample*.

6.4. Preparation of final samples

The material in each aggregate sample shall be carefully mixed to obtain an homogenized sample (e). If necessary the aggregate sample should first be reduced to at least 2 kg or two litres (reduced sample) either by using a mechanical divider or by the quartering method.

At least three final samples shall then be prepared, of approximately the same amount and conforming to the quantitative requirements of 5.A.4 or 5.B.4. Each sample shall be put into an appropriate container. All necessary precautions shall be taken to avoid any change of composition of the sample, contamination or adulteration which might arise during transportation or storage.

(d) For packaged feedingstuffs, a part of the contents of the packages to be sampled shall be removed, using a spear or shovel, after having, if necessary, emptied the packages separately.

(e) Any lumps shall be broken up (if necessary by separating them out and returning them to the sample) in each aggregate sample separately.

6.5. **Packaging of final samples**

The containers of packages shall be sealed and labelled (the total label must be incorporated in the seal) in such a manner that they cannot be opened without damaging the seal.

7. **SAMPLING RECORD**

A record must be kept of each sampling, permitting each sampled portion to be identified unambiguously.

8. **DESTINATION OF SAMPLES**

For each aggregate sample, at least one final sample shall be sent as quickly as possible to the authorized analytical laboratory, together with the information necessary for the analyst.

SEVENTH COMMISSION DIRECTIVE

of 1 March 1976

establishing Community methods of analysis for the official control of feedingstuffs

(76/372/EEC)

THE COMMISSION OF THE EUROPEAN
COMMUNITIES,

HAS ADOPTED THIS DIRECTIVE:

Article 1

Having regard to the Treaty establishing the European Economic Community,

Having regard to the Council Directive of 20 July 1970 on the introduction of Community methods of sampling and analysis for the official control of feedingstuffs ⁽¹⁾, as last amended by the Act of Accession ⁽²⁾, and in particular Article 2 thereof,

Whereas that Directive requires that official control of feedingstuffs be carried out using Community methods of sampling and analysis for the purpose of checking compliance with requirements arising under the provisions laid down by law, regulation or administrative action concerning the quality and composition of feedingstuffs;

Whereas Commission Directives 71/250/EEC of 15 June 1971 ⁽³⁾, 71/393/EEC of 18 November 1971 ⁽⁴⁾, 72/199/EEC of 27 April 1972 ⁽⁵⁾, 73/46/EEC of 5 December 1972 ⁽⁶⁾, 74/203/EEC of 25 March 1974 ⁽⁷⁾ and 75/84/EEC of 20 December 1974 ⁽⁸⁾ have already established a number of Community methods of analysis; whereas the progress of work since then makes it advisable to adopt a seventh set of methods;

Whereas the measures provided for in this Directive are in accordance with the opinion of the Standing Committee for Feedingstuffs,

The Member States shall require that analyses for official controls of feedingstuffs, as regards their content of aflatoxin B₁, be carried out in accordance with the methods described in the Annex to this Directive.

The general provisions set out in Part 1 (Introduction) of the Annex to first Commission Directive 71/250/EEC of 15 June 1971, with the exception of the part dealing with preparation of the sample to be analyzed, shall be applicable to the methods described in the Annex to this Directive.

Article 2

The Member States shall, not later than 1 October 1976, bring into force the laws, regulations or administrative provisions necessary to comply with this Directive. They shall forthwith notify the Commission thereof.

Article 3

This Directive is addressed to the Member States.

Done at Brussels, 1 March 1976.

For the Commission

P. J. LARDINOIS

Member of the Commission

⁽¹⁾ OJ No L 170, 3. 8. 1970, p. 2.

⁽²⁾ OJ No L 73, 27. 3. 1972, p. 14.

⁽³⁾ OJ No L 155, 12. 7. 1971, p. 13.

⁽⁴⁾ OJ No L 279, 20. 12. 1971, p. 7.

⁽⁵⁾ OJ No L 123, 29. 5. 1972, p. 6.

⁽⁶⁾ OJ No L 83, 30. 3. 1973, p. 21.

⁽⁷⁾ OJ No L 108, 22. 4. 1974, p. 7.

⁽⁸⁾ OJ No L 32, 5. 2. 1975, p. 26.

ANNEX

DETERMINATION OF AFLATOXIN B₁

A. ONE-DIMENSIONAL THIN LAYER CHROMATOGRAPHIC METHOD

1. Purpose and scope

The method makes it possible to determine the level of aflatoxin B₁ in the following feedingstuffs: groundnut, copra, linseed, soya, sesame, babassu palm and maize-germ oil cakes, cereals and cereal products, pea meal, potato pulp and starch. The lower limit of determination is 0.01 mg/kg (10 ppb).

If the presence of interfering substances hinders the determination, it is necessary to start the analysis again using method B (two-dimensional thin layer chromatography).

2. Principle

The sample is subjected to extraction with chloroform. The extract is filtered, and an aliquot portion taken and purified by column chromatography on silica gel. The eluate is evaporated and the residue redissolved in a specific volume of chloroform or of a mixture of benzene and acetonitrile. An aliquot portion of this solution is subjected to thin-layer chromatography (TLC). The quantity of aflatoxin B₁ is determined under UV irradiation of the chromatogram, either visually or by fluorodensitometry, by comparison with known quantities of standard aflatoxin B₁. The identity of the aflatoxin B₁ extracted from the feedingstuff must be confirmed by the procedure indicated.

3. Reagents

NB: All the reagents must be of 'analytical reagent' quality unless otherwise stated.

- 3.1. Acetone.
- 3.2. Chloroform, stabilized with 0.5 to 1.0 % of 96 % ethanol (v/v).
- 3.3. N-hexane.
- 3.4. Methanol.
- 3.5. Anhydrous diethyl ether, free from peroxides.
- 3.6. Mixture of benzene and acetonitrile: 98/2 (v/v).
- 3.7. Mixture of chloroform (3.2) and methanol (3.4) : 97/3 (v/v).
- 3.8. Silica gel, for column chromatography, particle size 0.05 to 0.20 mm.
- 3.9. Absorbent cotton wool, previously defatted with chloroform, or glass wool.
- 3.10. Sodium sulphate, anhydrous, granular.
- 3.11. Inert gas, e.g. nitrogen.
- 3.12. 1 N Hydrochloric acid.
- 3.13. 50 % (v/v) sulphuric acid.
- 3.14. Kieselguhr (hyfflosupercel), washed in acid.
- 3.15. Silica gel G-HR or equivalent, for TLC.
- 3.16. Standard solution with about 0.1 µg of aflatoxin B₁ per ml in chloroform (3.2) or the benzene/acetonitrile mixture (3.6), prepared and checked as indicated in Section 7.

- 3.17. Standard solution for qualitative testing purposes containing about 0.1 µg of aflatoxin B₁ and B₂ per ml in chloroform (3.2) or the benzene/acetonitrile mixture (3.6). These concentrations are given as a guide. They must be adjusted so as to obtain the same intensity of fluorescence for both aflatoxins.
- 3.18. Developing solvents:
- 3.18.1. Chloroform (3.2)/acetone (3.1): 9/1 (v/v), unsaturated tank;
- 3.18.2. Diethyl ether (3.5)/methanol (3.4)/water: 96/3/1 (v/v/v), unsaturated tank;
- 3.18.3. Diethyl ether (3.5)/methanol (3.4)/water: 94/4.5/1.5 (v/v/v), saturated tank;
- 3.18.4. Chloroform (3.2)/methanol (3.4): 94/6 (v/v), saturated tank;
- 3.18.5. Chloroform (3.2)/methanol (3.4): 97/3 (v/v), saturated tank.

4. Apparatus

- 4.1. Grinder-mixer.
- 4.2. Shaking apparatus or magnetic stirrer.
- 4.3. Fluted filter papers, Schleicher and Schüll No 588 or equivalent, diameter: 24 cm
- 4.4. Glass tube for chromatography (internal diameter: 22 mm, length: 300 mm), with a PTFE cock and a 250-ml reservoir.
- 4.5. Rotary vacuum evaporator, with a 500-ml round-bottom flask.
- 4.6. 500-ml conical flasks, with ground-glass stoppers.
- 4.7. TLC apparatus.
- 4.8. Glass plates for TLC, 200 × 200 mm, prepared as follows (the quantities indicated are sufficient to cover five plates). Put 30 g of silica gel G-HR (3.15) into a conical flask. Add 60 ml water, stopper and shake for a minute. Spread the suspension on the plates so as to obtain a uniform layer 0.25 mm thick. Leave to dry in the air and then store in a desiccator containing silica gel. At the time of use, activate the plates by keeping them in the oven at 110 °C for one hour.
- Ready-to-use plates are suitable if they give results similar to those obtained with the plates prepared as indicated above.
- 4.9. Long-wavelength (360 nm) UV lamp. The intensity of irradiation must make it possible for a spot of 1 ng of aflatoxin B₁ to be still clearly distinguished on a TLC plate at a distance of 10 cm from the lamp.
- 4.10. 10 ml graduated tubes with polyethylene stoppers.
- 4.11. UV spectrophotometer.
- 4.12. Fluorodensitometer (optional).

5. Procedure

- 5.1. *Preparation of the sample* (see under 'Observations', Part C, point 1).
- Grind the sample so that the whole of it will pass through a sieve with a 1-mm mesh (in accordance with recommendation ISO R 565).
- 5.2. *Extraction*
- Put 50 g of ground, homogenized sample into a 500-ml conical flask (4.6). Add 25 g of Kieselguhr (3.14), 25 ml of water and 250 ml of chloroform (3.2). Stopper the flask, shake or stir for 30 minutes with the apparatus (4.2) and filter through a fluted filter paper (4.3). Discard the first 10 ml of the filtrate and then collect 50 ml.

5.3. *Column clean-up*

Insert into the lower end of a chromatography tube (4.4) a cotton or glass wool plug (3.9), fill two-thirds of the tube with chloroform (3.2) and add 5 g of sodium sulphate (3.10).

Check that the upper surface of the sodium sulphate layer is flat, then add 10 g of silica gel (3.8) in small portions. Stir carefully after each addition to eliminate air bubbles. Leave to stand for 15 minutes and then carefully add 15 g of sodium sulphate (3.10). Let the liquid fall until it is just above the upper surface of the sodium sulphate layer.

Mix the 50 ml of extract collected in 5.2 with 100 ml of n-hexane (3.3) and quantitatively transfer the mixture to the column. Let the liquid fall until it is just above the upper surface of the sodium sulphate layer. Discard this washing. Then add 100 ml of diethylether (3.5) and again allow it to fall to the upper surface of the sodium sulphate layer. During these operations see that the rate of flow is 8 to 12 ml per minute and that the column does not run dry. Discard the liquid that comes out. Then elute with 150 ml of the chloroform/methanol mixture (3.7) and collect the whole of the eluate.

Evaporate the latter almost to dryness at a temperature not exceeding 50 °C and under a stream of inert gas (3.11) with the rotary evaporator (4.5). Quantitatively transfer the residue, using chloroform (3.2) or the benzene-acetonitrile mixture (3.6), to a 10 ml graduated tube (4.10). Concentrate the solution under a stream of inert gas (3.11) and then adjust the volume to 2 ml with chloroform (3.2) or the benzene/acetonitrile mixture (3.6).

5.4. *Thin-layer chromatography*

Spot on a TLC plate (4.8), 2 cm from the lower edge and at intervals of 2 cm, the volumes indicated below of the standard solution and the extract:

- 10, 15, 20, 30 and 40 μ l of the standard aflatoxin B₁ solution (3.16);
- 10 μ l of the extract obtained in 5.3 and, *superimposed on the same point*, 20 μ l of the standard solution (3.16);
- 10 and 20 μ l of the extract obtained in 5.3.

Develop the chromatogram in the dark with one of the developing solvents (3.18). The choice of the solvent must be made beforehand, by depositing 25 μ l of the qualitative standard solution (3.17) on the plate and checking that, when developed, aflatoxin B₁ and B₂ are completely separated.

Let the solvents evaporate in the dark and then irradiate with UV light, placing the plate 10 cm from the lamp (4.9). The spots of aflatoxin B₁ give a blue fluorescence.

5.5. *Quantitative determinations*

Determine either visually or by fluorodensitometry as indicated below.

5.5.1. *Visual measurements*

Determine the quantity of aflatoxin B₁ in the extract by matching the fluorescence intensity of the extract spots with that of one of the standard solution spots. Interpolate if necessary. The fluorescence obtained by the superimposition of the extract on the standard solution must be more intense than that of the 10 μ l of extract and there must not be more than one visible spot. If the fluorescence intensity given by the 10 μ l of extract is greater than that of the 40 μ l of standard solution, dilute the extract 10 or 100 times with chloroform (3.2) or the benzene/acetonitrile mixture (3.6) before subjecting it again to thin-layer chromatography.

5.5.2. *Measurements by fluorodensitometry*

Measure the fluorescence intensity of the aflatoxin B₁ spots with the fluorodensitometer (4.12) at an excitation wavelength of 365 nm and an emission wavelength of 443 nm. Determine the quantity of aflatoxin B₁ in the extract spots by comparison of their fluorescence intensities with that of the standard aflatoxin B₁ spots.

5.6. *Confirmation of the identity of aflatoxin B₁*

Confirm the identity of the aflatoxin B₁ in the extract by the processes indicated below.

5.6.1. Treatment with sulphuric acid

Spray sulphuric acid (3.13) on to the chromatogram obtained in 5.4. The fluorescence of the aflatoxin B₁ spots must turn from blue to yellow under UV irradiation.

5.6.2. Two-dimensional chromatography involving the formation of aflatoxin B₁-hemiacetal (aflatoxin B_{2a})

NB: The operations described below must be carried out following carefully the diagram in figure 3.

5.6.2.1. Application of the solutions

Score two straight lines on the plate (4.8) parallel to two contiguous sides (6 cm in from each side) to limit migration of the solvent fronts. Spot the following solutions on the plate using capillary pipettes or microsyringes:

- on point A: a volume of purified extract of the sample, obtained in 5.3, containing about 2.5 nm of aflatoxin B₁;
- on points B and C: 25 µl of the standard solution (3.16).

5.6.2.2. Development

Develop the chromatogram in direction I, in the dark, using the developing solvent (3.18.1) (1 cm layer in an unsaturated tank) until the solvent front reaches the solvent limit line.

Remove the plate from the tank and allow to dry in the dark at ambient temperature for five minutes. Then spray hydrochloric acid (3.12) along a band 2.5 cm high, covering points A and B (indicated by the hatched area in figure 3) until it darkens, protecting the rest of the plate with a glass sheet. Allow to react for 10 minutes in the dark and dry with a stream of air at ambient temperature.

Next, develop the chromatogram in direction II, in the dark, using the developing solvent (3.18.1) (1 cm layer in an unsaturated tank) until the solvent front reaches the solvent limit line. Remove the plate from the tank and allow to dry at ambient temperature.

5.6.2.3. Interpretation of the chromatogram

Examine the chromatogram under UV light (4.9) and check for the following features.

- (a) Appearance of a blue fluorescent spot of aflatoxin B₁ originating from the standard solution applied at C (migration in the direction I).
- (b) Appearance of a blue fluorescent spot of unreacted (with the hydrochloric acid) aflatoxin B₁ and a more intense blue fluorescent spot of aflatoxin B₁-hemiacetal, both originating from the standard solution applied at B (migration in direction II).
- (c) Appearance of spots matching those described in (b), originating from the sample extract applied at A. The position of these spots is defined first by the migration distance of the aflatoxin B₁ from point A in direction I (the same as that travelled by the standard applied at C), and then by the migration distances from there in direction II of the aflatoxin B₁-hemiacetal (same as those travelled by the standard applied at B). The fluorescence intensities of hemiacetal spots originating from the extract and from the standard applied at B should match.

6. Calculation of the results

6.1. From the visual measurements

The content in micrograms of aflatoxin B₁ per kg of sample (ppb) is given by the formula:

$$\frac{S \cdot Y \cdot V}{W \cdot X}$$

in which:

Y and X are respectively the volumes in microlitres of the standard solution of aflatoxin B₁ (3.16) and of the extract having a similar intensity of fluorescence;

S = concentration in micrograms of aflatoxin B₁ per ml in the standard solution (3.16);

V = final volume of the extract in microlitres, allowing for any dilution that was necessary;

W = weight in grammes of the sample corresponding to the volume of extract subjected to column clean-up.

6.2. *From the fluorodensitometric measurements*

The content in micrograms of aflatoxin B₁ per kg of sample is given by the formula:

$$\frac{S \cdot V}{W \cdot Y}$$

in which:

Y = volume in microlitres of the extract spotted on the plate (10 or 20 µl);

S = quantity in nanograms of aflatoxin B₁ in the extract spot (proportional to the value of Y taken), deduced from the measurements;

V = final volume of the extract in microlitres, allowing for any dilution that was necessary;

W = weight in grammes of the sample corresponding to the volume of extract subjected to column clean-up.

7. **Preparation and testing of the standard solution (3.16)**

7.1. *Determination of the concentration of aflatoxin B₁*

Prepare a standard solution of aflatoxin B₁ in the chloroform (3.2) or the benzene/acetonitrile mixture (3.6) with a concentration of 8 to 10 µg/ml. Determine the absorption spectrum between 330 and 370 nm with the aid of the spectrophotometer (4.11).

Measure the optical density (A) at 363 nm in the case of the chloroform solution; or at 348 nm in the case of the solution in the benzene/acetonitrile mixture.

Calculate the concentration in micrograms of aflatoxin B₁ per ml of solution from the formulae below:

$$\frac{312 \cdot A \cdot 1000}{20600} \text{ for the chloroform solution;}$$

$$\frac{312 \cdot A \cdot 1000}{19800} \text{ for the solution in the benzene/acetonitrile mixture.}$$

Dilute as appropriate, away from day light, to obtain a working standard solution with a concentration of aflatoxin B₁ of about 0.1 µg/ml. If kept in a refrigerator at 4 °C, this solution is stable for two weeks.

7.2. *Testing of chromatographic purity*

Spot on a plate (4.8) 5 µl of the standard solution of aflatoxin B₁ containing 8 to 10 µg/ml (7.1). Develop the chromatogram as indicated in 5.4. In UV light the chromatogram should show only one spot and no fluorescence must be perceptible in the original deposit zone.

8. **Repeatability**

The difference between the results of two parallel determinations carried out on the same sample by the same analyst should not exceed:

- 25 % related to the highest result for contents of aflatoxin B₁ from 10 and up to 20 µg/kg;
- 5 µg, in absolute value, for contents greater than 20 and up to 50 µg/kg;
- 10 % related to the highest value for contents above 50 µg/kg.

9. **Reproducibility**

See under 'Observations', Part C, point 2.

B TWO-DIMENSIONAL THIN LAYER CHROMATOGRAPHIC METHOD**1. Purpose and scope**

The method makes it possible to determine the level of aflatoxin B₁ in feedingstuffs not falling within the scope of method A. The lower limit of determination is 0.01 mg/kg (10 ppb). The method is not applicable to feedingstuffs containing citrus pulp.

2. Principle

The sample is subjected to extraction with chloroform. The extract is filtered, and an aliquot portion taken and purified by column chromatography on silica gel. The eluate is evaporated and the residue redissolved in a specific volume of chloroform or of a mixture of benzene and acetonitrile. An aliquot portion of this solution is subjected to two-dimensional thin-layer chromatography (TLC). The quantity of aflatoxin B₁ is determined under UV irradiation of the chromatogram, either visually or by fluorodensitometry, by comparison with known quantities of standard aflatoxin B₁. The identity of the aflatoxin B₁ extracted from the feedingstuff must be confirmed by the procedure indicated.

3. Reagents

NB: All the reagents must be of 'analytical reagent' quality unless otherwise stated.

- 3.1. Acetone.
- 3.2. Chloroform, stabilized with 0.5 to 1 % of 96 % ethanol (v/v).
- 3.3. N-hexane.
- 3.4. Methanol.
- 3.5. Anhydrous diethyl ether, free from peroxides.
- 3.6. Mixture of benzene and acetonitrile: 98/2 (v/v).
- 3.7. Mixture of chloroform (3.2) and methanol (3.4): 97/3 (v/v).
- 3.8. Silica gel, for column chromatography, particle size 0.05 to 0.20 mm.
- 3.9. Absorbent cotton wool, previously defatted with chloroform, or glass wool.
- 3.10. Sodium sulphate, anhydrous, granular.
- 3.11. Inert gas, e.g. nitrogen.
- 3.12. 1 n-hydrochloric acid.
- 3.13. Kieselguhr, (hyflo-supercel), washed in acid.
- 3.14. Silica gel G-HR or equivalent, for TLC.
- 3.15. Developing solvents.
 - 3.15.1. Diethyl ether (3.5)/methanol (3.4)/water: 94/4.5/1.5 (v/v/v), saturated tank.
 - 3.15.2. Chloroform (3.2)/acetone (3.1): 9/1 (v/v), unsaturated tank.
- 3.16. Standard solution with about 0.1 µg aflatoxin B₁ per ml in chloroform (3.2) or the benzene/acetonitrile mixture (3.6), prepared and checked as described in point 7 of method A.

4. Apparatus

See under point 4 of method A.

5. Procedure

- 5.1. *Preparation of the sample*
5.2. *Extraction*
5.3. *Column clean-up*
- } See under points 5.1, 5.2 and 5.3 of method A.

5.4. Two dimensional TLC

5.4.1. Application of the solutions (follow the diagram in figure 1)

Score two straight lines on a plate (4.8) parallel to two contiguous sides (5 and 6 cm from each side respectively), to limit migration of the solvent fronts. Spot the following solutions on the plate using capillary pipettes or micro-syringes:

- on point A, 20 μ l of the purified sample extract obtained in 5.3;
- on point B, 20 μ l of the standard solution (3.16);
- on point C, 10 μ l of the standard solution (3.16);
- on point D, 20 μ l of the standard solution (3.16);
- on point E, 40 μ l of the standard solution (3.16).

Dry in a slow stream of air or inert gas (3.11). The spots obtained must have a diameter of about 5 mm.

5.4.2. Development (follow the diagram in figure 1)

Develop the chromatogram in direction I, in the dark, using the developing solvent (3.15.1) (1-cm layer in a saturated tank) until the solvent front reaches the solvent limit line. Remove the plate from the tank and allow to dry in the dark at ambient temperature for 15 minutes.

Then develop the chromatogram in direction II, in the dark, using the developing solvent (3.15.2) (1-cm layer in an unsaturated tank) until the solvent front reaches the solvent limit line. Remove the plate from the tank and allow to dry, in the dark, at ambient temperature.

5.4.3. Interpretation of the chromatogram (follow the diagram in figure 2)

Irradiate the chromatogram with UV light by placing the plate 10 cm from the (4.9). Locate the position of the blue fluorescent spots B, C, D and E of the aflatoxin B₁ from the standard solution. Project two imaginary lines passing through these spots and at right angles to the development directions. The intersection P of these lines is the location in which to expect to find the aflatoxin B₁ spot originating from the sample extract applied at A (figure 1). However, the actual location of the aflatoxin B₁ spot may be at a point Q at the intersection of two imaginary straight lines forming an angle of about 100° between them and passing through spots B and C respectively. Determine the quantity of aflatoxin B₁ in the sample extract as indicated in 5.5.

5.4.4. Supplementary chromatography

Score two straight lines on a new plate (4.8) parallel to two contiguous sides, as indicated in the diagram in figure 1, and apply on point A (see figure 1) 20 μ l of the purified sample extract obtained in 5.3 and, superimposed on it, 20 μ l of the standard solution (3.16). Develop as indicated in 5.4.2. Irradiate the chromatogram with UV light (4.9) and check that:

- the aflatoxin B₁ spots from the extract and the standard solution are superimposed,
- the fluorescence of this spot is more intense than that of the aflatoxin B₁ spot developed at Q on the first plate.

5.5. Quantitative determinations

Determine either visually or by fluorodensitometry as indicated below.

5.5.1. Visual measurements

Determine the quantity of aflatoxin B₁ in the extract by matching the fluorescence intensity of the extract spot with that of spots C, D and E of the standard solution. Interpolate if necessary. If the fluorescence intensity given by the 20 μ l of extract

is greater than that of the 40 μ l of standard solution, dilute the extract 10 or 100 times with chloroform (3.2) or the benzene/acetonitrile mixture (3.6) before subjecting it again to thin-layer chromatography.

5.5.2. Measurements by fluorodensitometry

Measure the fluorescence intensity of the aflatoxin B₁ spots with the fluorodensitometer (4.12), using an excitation wavelength of 365 nm and an emission wavelength of 443 nm.

Determine the quantity of aflatoxin B₁ in the extract spot by comparison of its fluorescence intensity with that of spots C, D and E of the standard solution.

5.6. Confirmation of the identity of aflatoxin B₁

See under point 5.6 of method A.

6. Calculation of the results

See under point 6 of method A.

7. Repeatability

See under point 8 of method A.

8. Reproducibility

See under 'Observations', Part C, point 2.

C. OBSERVATIONS CONCERNING METHODS A AND B

1. Defatting

Samples containing more than 5% fats must be defatted with light petroleum (bp 40 to 60 °C) after the preparation indicated in 5.1.

In such cases, the analytical results must be expressed in terms of the weight of the non-defatted sample.

2. Reproducibility of the results

The reproducibility of the results, i.e. the variation between the results obtained by two or more laboratories on the same sample has been estimated at:

± 50% of the mean value for mean values of aflatoxin B₁ from 10 and up to 20 μ g/kg;

± 10 μ g/kg on the mean value for mean values greater than 20 and up to 50 μ g/kg;

± 20% of the mean value for mean values above 50 μ g/kg.

Appendix to Annex

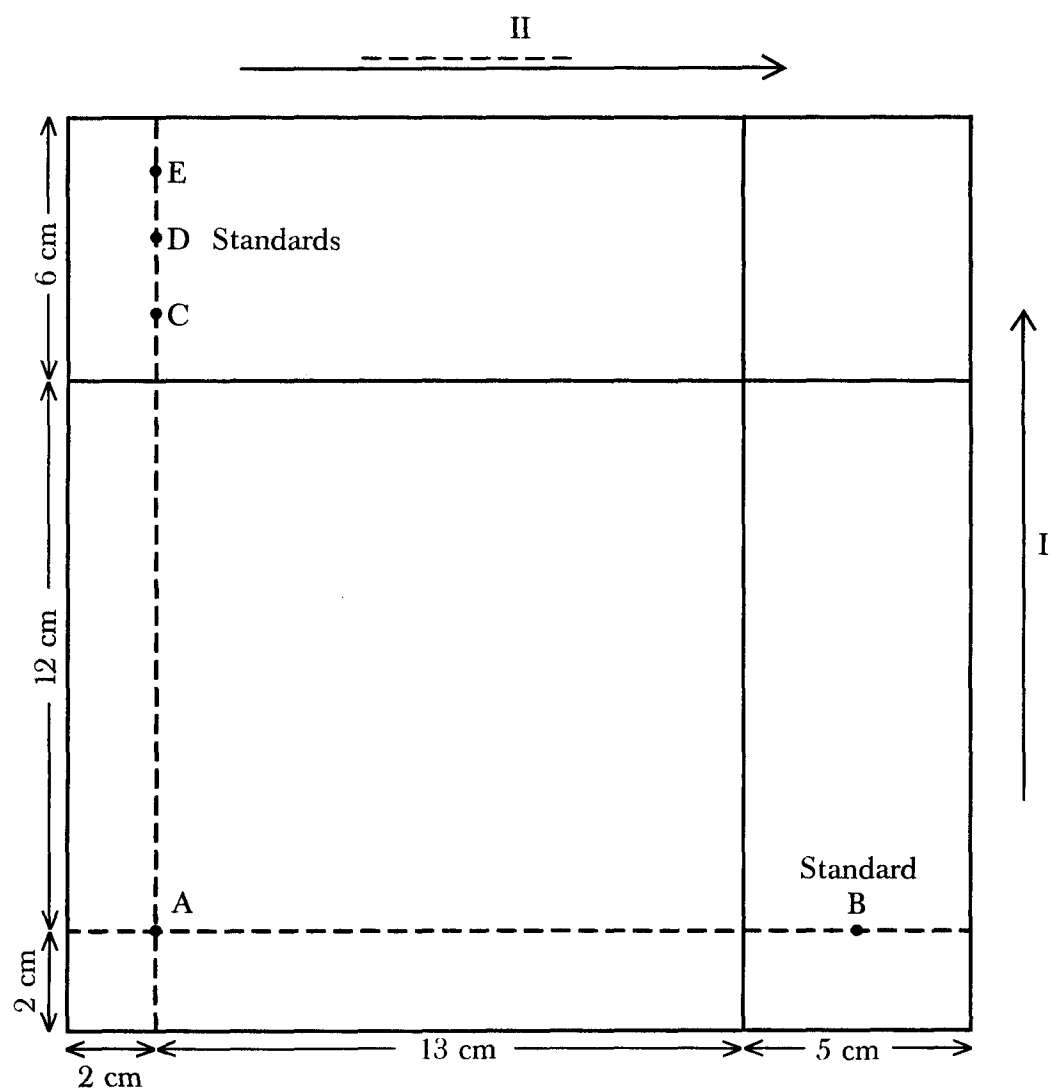


Figure 1

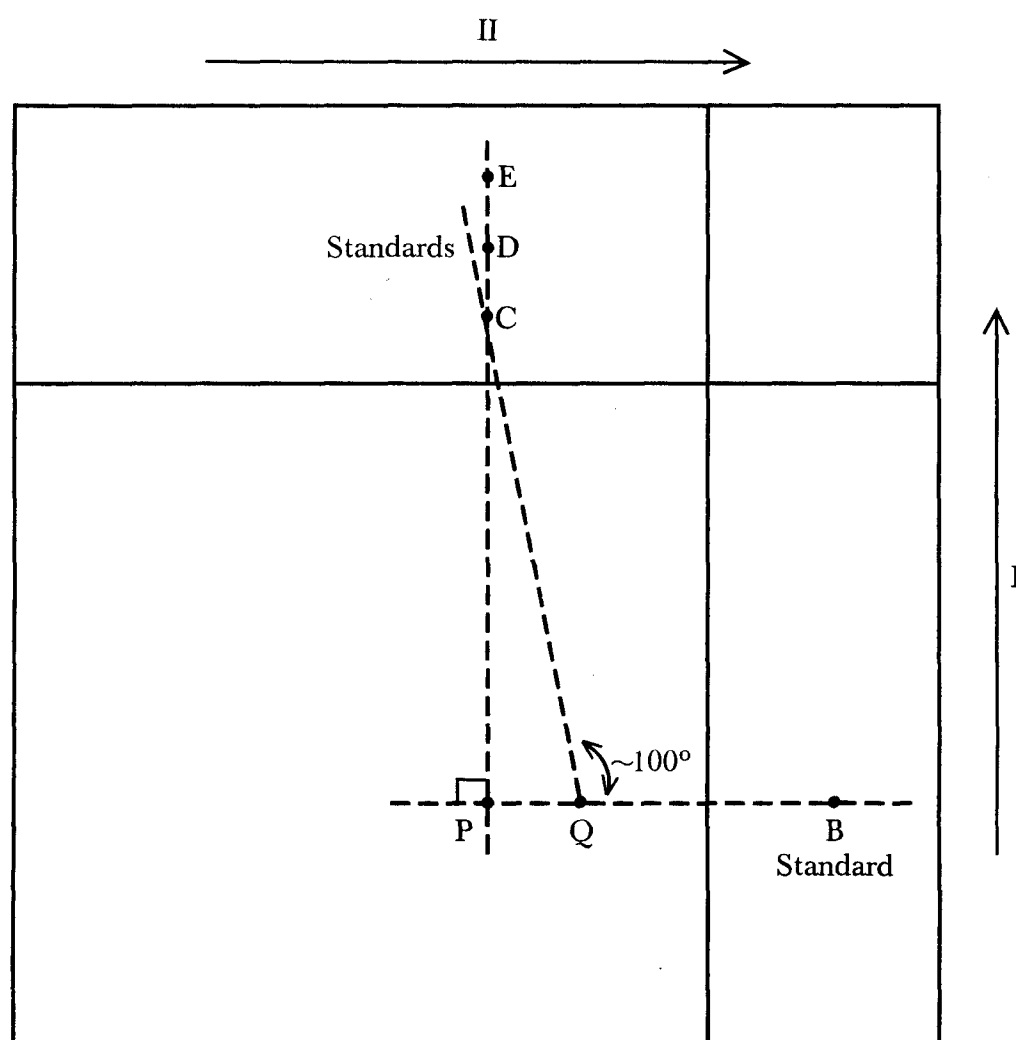
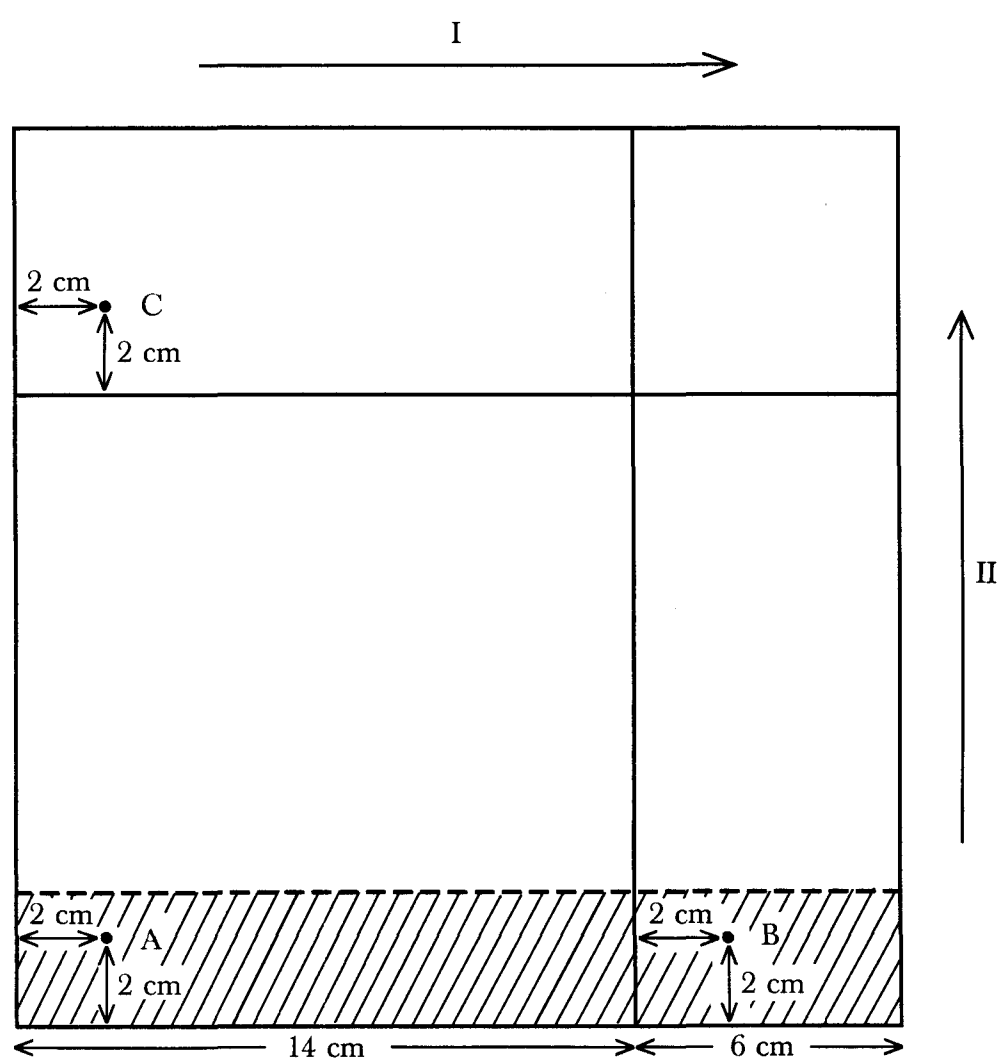


Figure 2

*Figure 3*

COMMISSION DECISION

of 3 March 1976

on the implementation of the reform of agricultural structures in the United Kingdom
pursuant to Title II of Directive 75/268/EEC

(Only the English text is authentic)

(76/373/EEC)

THE COMMISSION OF THE EUROPEAN
COMMUNITIES,

Having regard to the Treaty establishing the
European Economic Community,

Having regard to Council Directive 75/268/EEC of
28 April 1975 on mountain and hill farming and
farming in certain less-favoured areas ⁽¹⁾, and in
particular Article 13 thereof,

Having regard to Council Directive 72/159/EEC of
17 April 1972 on the modernization of farms ⁽²⁾, and
in particular Article 18 (3) thereof,

Whereas on 22 October 1975 the Government of
the United Kingdom forwarded, pursuant to
Article 13 of Directive 75/268/EEC, in conjunction
with the second indent of Article 17 (1) of Directive
72/159/EEC, the following statutory instruments
promulgated by the Minister of Agriculture, Fisheries
and Food:

England and Wales:

1972 No 1616, 1972 No 1617, 1974 No 1794, 1975
No 1140, 1973 No 828, 1973 No 829, 1974 No 1997;

Scotland:

1972 No 1659, 1972 No 1660, 1974 No 1863,
1975 No 1418, 1973 No 866, 1973 No 867, 1974
No 1906;

Northern Ireland:

1972 No 1618, 1972 No 1619, 1974 No 1795, 1975
No 1141, 1973 No 830, 1973 No 831, 1974 No 1996;

Whereas these statutory instruments, which were in
application before Directive 75/268/EEC came into
force, provide for the granting of subsidies on
breeding cows and sheep maintained in less-favoured
areas listed in Directive 75/276/EEC;

Whereas on 6 November 1975 the Government of
the United Kingdom also forwarded the draft of
a statutory instrument on stock breeding on hill
land (compensatory allowance) 1975 which will
replace the statutory instruments listed above with
effect from 1 January 1976; whereas the Commission
has, pursuant to Article 17 (3) of Directive
72/159/EEC, stated in its opinion of 30 January
1976 that this draft satisfies the conditions for
financial contribution by the Community;

Whereas, under Article 18 (3) of Directive
72/159/EEC and Article 13 of Directive
75/268/EEC, the Commission must decide whether,
having regard to the objectives of Directive
75/268/EEC and to the need for a proper connection
between the various measures, the laws, regulations
or administrative provisions forwarded comply with
this Directive and thus satisfy the conditions for
financial contribution by the Community to common
measures undertaken in 1975;

Whereas the basic objective of Directive 75/268/EEC
is to ensure the continuation of farming in the
mountain and hill areas and less-favoured areas
laid down by the Council, thereby maintaining a
minimum population level or conserving the
countryside;

Whereas Directive 75/268/EEC therefore authorizes
Member States to introduce a system of aids to
encourage farming and to raise farm incomes in
these areas;

Whereas this system may take the form of an
allowance to compensate farmers for permanent
natural handicaps, if they undertake to pursue a
farming activity in accordance with the aims of the
Directive for at least five years; whereas in the case
of cattle, sheep and goat farming this allowance is
calculated in relation to livestock numbers and may
not exceed 50 units of account per livestock unit
and 50 units of account per hectare under forage,
but may not be less than 15 units of account per
livestock unit; whereas in addition to the conditions
in Articles 6 and 7 of the Directive, the Member

⁽¹⁾ OJ No L 128, 19. 5. 1975, p. 1.

⁽²⁾ OJ No L 96, 23. 4. 1972, p. 1.

States may lay down supplementary conditions or limitations on the granting of compensatory allowances;

Whereas the abovementioned statutory instruments already essentially comply with the aims and conditions of Directive 75/268/EEC;

Whereas, however they fail to provide that farmers receiving the compensatory allowance should give the undertaking referred to in Article 6 (1) of Directive 75/268/EEC to pursue a farming activity for at least five years; whereas, in addition, for England and Wales and for Northern Ireland they also permit the compensatory allowance in a few cases to be slightly higher than 50 units of account per hectare under forage;

Whereas, pursuant to the second sentence of Article 5 and Article 17 of Directive 75/268/EEC, the abovementioned statutory instruments must, on both these points, be amended to comply with this Directive within one year from the date on which the Directive came into effect; whereas the abovementioned draft statutory instrument on stock breeding on hill land (compensatory allowances) 1975 already provides for the corresponding amendments;

Whereas, so far as the undertaking in Article 6 (1) of Directive 75/268/EEC is concerned, it is possible to restrict the financial contribution of the Community to those cases where the recipients of the subsidies give such an undertaking on the occasion of the granting of compensatory allowances in 1976;

Whereas, so far as the possible excess of the maximum amount of the compensatory allowance over that laid down in Article 7 (1) of Directive 75/268/EEC of 50 units of account per hectare is concerned in England and Wales and Northern Ireland, such excess is very insignificant;

Whereas therefore subject to the necessary adjustments being made in 1976, although this excess is not compatible with the Directive, a financial contribution by the Community to the cost of the statutory instruments referred to above for 1975 can be held to be justified, bearing in mind the transitional period allowed by Article 17 of the Directive;

Whereas the EAGGF Committee has been consulted on the financial aspects;

Whereas the measures provided for in this Decision are in accordance with the opinion of the Standing Committee on Agricultural Structures,

HAS ADOPTED THIS DECISION:

Article 1

The statutory instruments forwarded by the Government of the United Kingdom on 22 October 1975 and referred to individually in the preamble satisfy the conditions for a financial contribution by the Community in 1975 to the measures referred to in Article 13 of Directive 75/268/EEC and Article 15 of Directive 72/159/EEC.

Article 2

The EAGGF Guidance Section shall contribute to the financing of the following subsidies:

- those which were granted in 1975 to farmers who, on the occasion of the granting of a compensatory allowance in 1976 give the undertaking in Article 6 (1) of Directive 75/268/EEC;
- those which in England and Wales and Northern Ireland do not exceed the maximum amount of 50 units of account per livestock unit and per hectare under forage permitted by Article 7 (1) of Directive 75/268/EEC.

Article 3

This Decision shall enter into force on the same day as the Commission Decision on the measures applied in the United Kingdom in 1976 to implement Title II of Directive 75/268/EEC.

Article 4

This Decision is addressed to the United Kingdom.

Done at Brussels, 3 March 1976.

For the Commission

P. J. LARDINOIS

Member of the Commission

COMMISSION DECISION

of 3 March 1976

on the implementation of the reform of agricultural structures in the Federal Republic of Germany pursuant to Title II of Directive 75/268/EEC

(Only the German text is authentic)

(76/374/EEC)

THE COMMISSION OF THE EUROPEAN COMMUNITIES,

Having regard to the Treaty establishing the European Economic Community,

Having regard to Council Directive 75/268/EEC of 28 April 1975 on mountain and hill farming and farming, in certain less-favoured areas ⁽¹⁾, and in particular Article 13 thereof,

Having regard to Council Directive 72/159/EEC of 17 April 1972 on the modernization of farms ⁽²⁾, and in particular Article 18 (3) thereof,

Whereas on 26 June 1975 the Government of the Federal Republic of Germany forwarded, pursuant to Article 13 of Directive 75/268/EEC, principles for incentives to farms in mountain and hill areas and in certain less-favoured areas promulgated on 11 April 1975;

Whereas, on 11 November and 18 December 1975, the Government of the Federal Republic of Germany also forwarded, pursuant to Article 13 of Directive 75/268/EEC, the text of the following laws, regulations and administrative provisions of the German Länder:

Bavaria

Directives on the Bavarian grassland and hill programme of 14 June 1975, Section 2: 'Premiums for extensive cattle farming'.

Baden-Württemberg

Order to encourage to agricultural measures for conservation of cultivated areas (subsidies to sheep farms) of 8 July 1971, in the version of 2 August 1973.

Hesse

Directives to encourage measures for preservation of the countryside by farms in the version of 11 March 1975, Section 3 (3): 'Incentives for keeping breeding cows', Section 3 (4): 'Incentives for keeping store cattle' and Section 3 (5): 'Incentives for sheep farming'.

North-Rhine-Westphalia

Order of 27 April 1973, in the version of 1975, Section 3 (1) (1): 'Premiums for keeping breeding cows'.

Whereas, under Article 18 (3) of Directive 72/159/EEC and Article 13 of Directive 75/268/EEC, the Commission must decide whether, having regard to the objectives of Directive 75/268/EEC and to the need for a proper connection between the various measures, the laws, regulations and administrative provisions communicated comply with this Directive and thus satisfy the conditions for financial contribution by the Community;

Whereas the essential objective of Directive 75/268/EEC is to ensure the continuation of farming, in the mountain and hill areas and less-favoured areas laid down by the Council, thereby maintaining a minimum population level or conserving the countryside;

Whereas Directive 75/268/EEC thereof authorized Member States to introduce a system of aids to encourage farming and to raise farm incomes in these areas;

Whereas this system may take the form of an allowance to compensate farmers for permanent natural handicaps, if they undertake to pursue a farming activity in accordance with the aims of the Directive for at least five years; whereas in the case of cattle, sheep and goats this allowance is calculated in relation to livestock numbers and may not exceed 50 units of account per livestock unit and 50 units of account per hectare under forage, but may not be less than 15 units of account per livestock unit;

⁽¹⁾ OJ No L 128, 19. 5. 1975, p. 1.

⁽²⁾ OJ No L 96, 23. 4. 1972, p. 1.

whereas in addition to the conditions in Articles 6 and 7 of the Directive, Member States may lay down supplementary conditions or limitations on the granting of compensatory allowances;

Whereas the measures provided for in the abovementioned principles for incentives to farms in mountain and hill areas and certain less-favoured areas, promulgated on 11 April 1975, and in Section 2 of the abovementioned directive of the Land Bavaria, comply with the aim and conditions of Directive 75/268/EEC;

Whereas the abovementioned measures of the Länder Baden-Württemberg, Hesse and North-Rhine-Westphalia, which were introduced before the entry into force of Directive 75/268/EEC, do not on the other hand comply with the conditions of the Directive and must be repealed or amended within one year from the entry into force of the Directive, pursuant to Article 5, second paragraph, and Article 17 of the Directive;

Whereas however, these measures, which are in addition to the compensatory allowances provided for in the principles for incentives to farms in mountain and hill areas and certain less-favoured areas promulgated on 11 April 1975 and basically only apply in less-favoured areas in which the compensatory allowance is not granted, are, when taken together, only of slight importance; whereas therefore, despite the measures which are incompatible with the Directive, a financial contribution by the Community to the cost of the principles for incentives to farms in mountain and hill areas and in certain less-favoured areas promulgated on 11 April 1975, and to the abovementioned directives of Land Bavaria, can be held to be justified even during the transitional period allowed by Article 17;

Whereas the EAGGF Committee has been consulted on the financial aspects;

Whereas the measures provided for in this Decision are in accordance with the opinion of the Standing Committee on Agricultural Structures,

HAS ADOPTED THIS DECISION:

Article 1

The principles for incentives to farms in mountain and hill areas and in certain less-favoured areas promulgated on 11 April 1975, as forwarded by the Government of the Federal Republic of Germany, and the measures provided for in Section 2 of the directives of Land Bavaria on the Bavarian grassland and hill programme of 14 June 1975 also forwarded, comply with the conditions for financial contribution by the Community to the measures referred to in Article 13 of Directive 75/268/EEC and Article 15 of Directive 72/159/EEC.

Article 2

This Decision shall cease to have effect on 28 April 1976, unless by when the measures of Länder Baden-Württemberg, Hesse and North-Rhine-Westphalia, which are referred to in the preamble, have been amended, to comply with the conditions of Directive 75/268/EEC, or repealed.

Article 3

This Decision is addressed to the Federal Republic of Germany.

Done at Brussels, 3 March 1976.

For the Commission

P. J. LARDINOIS

Member of the Commission

COMMISSION DECISION

of 3 March 1976

on the implementation of the reform of agricultural structures in France pursuant to
Title II of Directive 72/161/EEC

(Only the French text is authentic)

(76/375/EEC)

THE COMMISSION OF THE EUROPEAN
COMMUNITIES,

Having regard to the Treaty establishing the
European Economic Community,

Having regard to Council Directive 72/161/EEC of
17 April 1972 concerning the provision of socio-
economic guidance for and the acquisition of
occupational skills by persons engaged in
agriculture ⁽¹⁾, and in particular Article 11 (3) thereof,

Whereas on 14 November 1975 the French
Government, pursuant to Article 10 (3) of Directive
72/161/EEC, forwarded the following list of courses
organized in France for the training and advanced
training of persons engaged in agriculture:

- (a) courses of 120 hours to four to five months for
the advanced training of persons who have
already attended course or have undergone
formal agricultural training attested by a diploma
or certificate;
- (b) courses of at least 200 hours for persons who,
in order to be eligible for certain assistance,
require to produce evidence that they have
attained a given level of occupational competence;
- (c) courses of from 800 to 1 200 hours leading to
the award of a specific proficiency certificate,
designed for the classic training of persons
engaged in agriculture who have not received
adequate agricultural training;
- (d) short courses of from 20 to 120 hours, the costs
of which are met by the 'Fonds d'assurance
formation';

Whereas the French Government also forwarded the
following laws, regulations and administrative
provisions, on the basis of which the above training
courses have been organized:

— Law No 71-575 of 16 July 1971 on organization
of advanced vocational training within established
further education;

— Decree No 71-978 of 10 December 1971 relating
to the 'Fonds d'assurance formation' (fund for
the provision of training);

— Decree Nos 71-980 and 71-981 of 10 December
1971 relating to financial assistance for persons
undergoing vocational training;

— Decree No 67-996 of 15 November 1967 relating
to types of agreement governing vocational
training;

— order of 27 April 1973 relating to the grant of
an installation allowance for young farmers in
certain regions, and circulars of 22 May and
30 August 1973;

Whereas, under Article 11 (3) of Directive
72/161/EEC, the Commission must decide whether,
having regard to the objectives of the said Directive
and to the need for a proper connection between
the various measures, the provisions thus forwarded
satisfy the conditions for financial contribution by
the Community;

Whereas the basic objective of Title II of Directive
72/161/EEC is to enable persons of 18 years and
over engaged in agriculture to acquire new
agricultural skills, or to improve those which they
already possess, so that they can integrate into
modern agriculture;

Whereas to achieve this aim, Member States are
obliged, in pursuance of Articles 5 (1) and 6 (1) of
Directive 72/161/EEC, in addition to the normal
agricultural courses of study organized as part of
the national education system to implement measures
to give farmers and hired and family agricultural
workers further training of a general, technical and
economic nature;

⁽¹⁾ OJ No L 96, 23. 4. 1972, p. 15.

Whereas, in accordance with the third indent of Article 12 (2) of Directive 72/161/EEC, the Guidance Section of the EAGGF, refunds to Member States, 25% of the expenditure incurred in connection with the measures, up to a maximum of 1 500 units of account for each person engaged in agriculture having completed a course of vocational training or advanced vocational training;

Whereas the training courses referred to in (a) to (c) above are compatible with the aim of Title II of the said Directive as defined above and satisfy the requirement to be met by full courses designed to enable persons engaged in agriculture to acquire new agricultural skills or to improve significantly those which they already possess;

Whereas the short training courses of 20 to 120 hours referred to in (d) above can, in principle, be considered as advanced training courses within the meaning of Article 5 (1) and of the third indent of Article 12 (2) of the said Directive; whereas, however, on account of the brevity of the training courses, isolated participation in one such course cannot be considered as constituting attendance at a full course of advanced vocational training within the meaning of the third indent of Article 12 (2) of the Directive;

Whereas the EAGGF Committee has been consulted on the financial aspects;

Whereas the measures provided for in this Decision are in accordance with the opinion of the Standing Committee on Agriculture Structures,

HAS ADOPTED THIS DECISION:

Article 1

The training and advanced training courses for persons engaged in agriculture on the basis of the laws, regulations and administrative provisions, the texts of which were forwarded by the French Government on 14 November 1975, and which are individually listed in the preamble, satisfy the conditions for financial contribution by the Community to common measures within the meaning of Article 8 of Directive 72/161/EEC.

The Guidance Section of the EAGGF shall refund costs in accordance with the third indent of Article 12 (2) of Directive 72/161/EEC for the short training courses referred to in the communication of 14 November 1975 from the French Government only in respect of those farmers who have attended more than one such course.

Article 2

This Decision is addressed to the French Republic.

Done at Brussels, 3 March 1976.

For the Commission

P. J. LARDINOIS

Member of the Commission

COMMISSION DECISION

of 16 March 1976

on the implementation of the reform of agricultural structures in Ireland pursuant to
Council Directive 72/159/EEC

(Only the English text is authentic)

(76/376/EEC)

THE COMMISSION OF THE EUROPEAN
COMMUNITIES,Having regard to the Treaty establishing the
European Economic Community,Having regard to Council Directive 72/159/EEC of
17 April 1972 on the modernization of farms ⁽¹⁾,
and in particular Article 18 (3) thereof,Whereas on 2 December 1975 the Government of
Ireland notified certain provisions fixing anew for
1975 the comparable earned income per labour unit
and indicating its annual rate of growth;Whereas under Article 18 (3) of Directive
72/159/EEC, the Commission must decide whether,
having regard to the abovementioned communi-
cation, the existing provisions in Ireland for the
implementation of Directive 72/159/EEC, which form
the subject of Commission Decision 75/100/EEC of
20 January 1975 on the reform of agricultural
structures to be effected in Ireland in implementation
of Directives 72/159/EEC and 72/160/EEC ⁽²⁾,
continue to satisfy the conditions for financial
contribution from the Community to common
measures as referred to in Article 15 of Directive
72/159/EEC;Whereas the comparable earned income per labour
unit for 1975 and its annual rate of growth fixed
in the abovementioned provisions satisfy the
objectives of Article 4 of Directive 72/159/EEC;Whereas the measures provided for in this Decision
are in accordance with the opinion of the Standing
Committee on Agricultural Structures,

HAS ADOPTED THIS DECISION:

Article 1

The provisions for the implementation of Directive 72/159/EEC, which were notified by the Government of Ireland on 19 September 1974, having regard to the provisions which were notified on 2 December 1975 fixing the comparable earned income per labour unit for 1975 and its annual rate of growth, continue to satisfy the conditions for financial contribution from the Community to common measures as referred to in Article 15 of the said Directive.

Article 2

This Decision is addressed to Ireland.

Done at Brussels, 16 March 1976.

For the Commission

P. J. LARDINOIS

Member of the Commission

⁽¹⁾ OJ No L 96, 23. 4. 1972, p. 1.

⁽²⁾ OJ No L 40, 14. 2. 1975, p. 61.